# Inhibition of JNK provides neuroprotection against 6-OHDA toxicity in Parkinson's disease in vitro model

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## INTRODUCTION

Parkinson's disease (PD) is a neurodegenerative disorder marked by death of dopaminergic neurons.
PD is caused by oxidative stress, which may be induced by pro-oxidants like 6-OHDA.
JNK is a major pro-apoptotic kinase, which plays a key role in dopaminergic degeneration.

### **OBJECTIVES**

The present study aimed to investigate the effect of pharmacological JNK inhibition in



**Fig. 4.** The effect of pre-treatment with JNK V on the viability of differentiated SH-SY5Y cells exposed to neurotoxin 6-OHDA. The data are presented as mean ± SD. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 vs 6-OHDA. C – control, untreated cells; JNK – JNK V.

#### cellular model for PD.

## **METHODS**

- 1. SH-SY5Y cells were differentiated with 10  $\mu$ M retinoic acid (RA) for 7 days prior to experiments.
- Neurodegeneration was induced by treatment with 6-OHDA at EC50. Cells were treated with JNK inhibitor V either 1 h before or after 6-OHDA-induced damage.
- 3. The effect of JNK V on cell viability upon 6-OHDA toxicity was measured by XTT assay.
- 4. The genotoxicity was assessed by the comet assay.
- 5. The mRNA expression level of specific pro-apoptotic ER stress-related genes was measured by RT-qPCR.

## RESULTS



**Fig. 1.** Morphological features of undifferentiated SH-SY5Y cells (**A**) and SH-SY5Y cells differentiated with RA (**B**). Undifferentiated cells are characterized by short processes and tend to grow in clusters, whereas differentiated cells have significantly longer neurites and communicate with each other via neural network.



**Fig. 2.** Cytotoxicity analysis of the used compounds' solvents in differentiated SH-SY5Y cells using XTT assay. The data are presented as mean ± SD. \*\*\*p<0.001 vs. NC. NC – negative control, untreated cells; DMSO – 0.1% DMSO; DPBS/AA – 0.5% DPBS with 0.15% w/v ascorbic acid, PC – positive control, cells treated with 20% DMSO.





**Fig. 5.** The effect of post-treatment with JNK V on the viability of differentiated SH-SY5Y cells exposed to neurotoxin 6-OHDA. The data are presented as mean ± SD. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 vs 6-OHDA. C – control, untreated cells; JNK – JNK V.

**Fig. 6.** Genotoxicity of the solvents and JNK V inhibitor in differentiated SH-SY5Y cells, and the effect of treatment with JNK V on the level of DNA damage in differentiated SH-SY5Y cells exposed to 6-OHDA. \*\*\*p<0.001 vs 6-OHDA. NC – negative control, untreated cells; DMSO – 0.1% DMSO; DPBS/AA – 0.5% DPBS with 0.15% ascorbic acid, PC – positive control, cells treated with 20% DMSO; JNK - JNK V.



**Fig. 7.** The mRNA expression levels of *MAPK10*, *XBP1* and *DDIT3* genes in differentiated SH-SY5Y cells exposed to 6-OHDA and treated with JNK V inhibitor. *GAPDH* was used as a reference gene. The data are presented as mean ± SD. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 vs 6-OHDA. C – control, untreated cells; JNK – JNK V.



**Fig. 3.** Cytotoxicity analysis of JNK V inhibitor in differentiated SH-SY5Y cells using XTT assay. The data are presented as mean ± SD. \*p<0.05, \*\*p<0.01, and \*\*\*p<0.001 vs. NC. NC – negative control, untreated cells; DMSO – 0.1% DMSO; PC – positive control, cells treated with 20% DMSO.

#### CONCLUSIONS

Small-molecule JNK V inhibitor is effective against 6-OHDA-induced damage *in vitro*. Thus, JNK inhibitors could potentially be applied for the selective treatment of PD.

This work was supported by grant PRELUDIUM BIS 3 no. 2021/43/O/NZ5/02068 from the National Science Centre, Poland.





